WHO Prequalification of In Vitro Diagnostics PUBLIC REPORT

Product: First Response Malaria Ag. *P.f. / P.v.* Card Test WHO reference number: PQDx 0329-010-00

First Response Malaria Ag. *P.f./P.v.* **Card Test** with product codes **PI19FRC10s**, **PI19FRC25s**, **PI19FRC30**, **PI19FRC25**, **PI19FRC05** and **PI19FRC10**, manufactured by **Premier Medical Corporation Private Limited**, **Rest-of-World regulatory version**, was accepted for the WHO list of prequalified in vitro diagnostics and was listed on 04 December 2018.

Summary of WHO Prequalification Assessment for the First Response Malaria Ag. P.f. / P.v. Card Test

	Date	Outcome
Prequalification listing	04 December 2018	listed
Dossier assessment	07 September 2018	MR
Site inspection(s) of quality management system	22-24 September 2024	MR
Product performance evaluation	2018	MR

MR: Meets Requirements

Report amendments and/or product changes

This public report has since been amended. Amendments may have arisen because of changes to the prequalified product for which WHO has been notified and has undertaken a review. Amendments to the report are summarized in the following table, and details of each amendment are provided below.

Public report amendment	Summary of amendment	Date of report amendment
Version 2	The introduction of new suppliers for Alcohol Swab and Twist Lancet resulted in the change of labels.	11 March 2025
Version 3.0	 Change in the regulatory certification and labelling of the supplier for sterile lancet "Shandong Lianfa Medical Plastic Products Co., Ltd." Change in the label of the alcohol swab supplied by Medtrue Enterprises Co., Limited. 	10 September 25

3. Addition of pack sizes of 05 and 10 tests/pack.	
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Intended use

According to the claim of manufacturer "First Response Malaria Ag. P.f. / P.v. Card Test is intended to be performed by trained users (in either laboratory or point of care settings) as qualitative screening in vitro diagnostic test for detection of Plasmodium falciparum and P. vivax. The test is intended to be used with human whole blood specimens (capillary or venous blood). Venous blood with the anti-coagulants such as heparin, EDTA or Sodium citrate do not affect the test results. The kit is not intended to be used for screening of blood donors. The test is not automated and does not require any additional instrument".

Assay description

According to the claim of manufacturer, "First Response Malaria Ag. P.f. / P.v. Card Test is based on principle of immunochromatography in which nitrocellulose membrane is precoated with two monoclonal antibodies as two separate lines. One monoclonal antibody (test line P.v.) is P. vivax specific to lactate dehydrogenase (pLDH) and the other line (test line P.f.) consists of a monoclonal antibody specific to histidine-rich protein 2 (HRP2) of the Plasmodium falciparum. When the test sample along with assay buffer flows through the nitrocellulose membrane, monoclonal antibodies conjugated with colloidal gold, which are P. vivax specific to pLDH and P. falciparum specific to HRP2 binds to Plasmodium antigens released from the lysed blood sample. These antigen-conjugated antibody complex moves through the nitrocellulose membrane and binds to corresponding immobilized antibody at test lines, which leads to the formation of colour line/lines indicating reactive results. The control line will appear irrespective of reactive or non-reactive sample.

The First Response® Malaria Ag. P.f./P.v. Card Test is "of additional value" in the differential diagnosis of Plasmodium falciparum and P. vivax."

Test kit contents:

Description	Configuration				
 Each single test pack contains: 1 × Test device & desiccant 1 × Specimen Transfer device 	10 × single test (product code PI19FRC10s)	1 × Master Instructions for Use			
 1 × Alcohol swab 1 × Sterile lancet 1 × Buffer vial 1 × Instructions for use 	25 × single test (product code PI19FRC25s)	1 × Master Instructions for Use			
 Test device with desiccant Specimen Transfer device 	30 × Bulk test (product code PI19FRC30)	n/a			
Buffer BottleSterile Lancet	25 × Bulk test (product code PI19FRC25)	n/a			

Alcohol swabs	05 × Bulk test	n/a
 Instructions for use 	(product code PI19FRC05)	,
	10 × Bulk test	n/2
	(product code PI19FRC10)	n/a

Items required but not provided:

- New pair of disposable gloves
- Permanent marker pen
- Timer
- Extra lancets and alcohol swabs if needed
- Sharp disposable box and biohazardous waste container
- Venipuncture blood collection materials and precision pipette plus tip (if whole blood is collected by venipuncture)
- Bio hazardous waste container

Storage

The test kit must be stored at 1 - 40 °C.

Shelf-life upon manufacture

24 months.

Warnings/limitations

Refer to the current version of the manufacturer's instructions for use.

Prioritization for pregualification

Based on the results of the WHO product testing of malaria RDTs for Round 8, First Response Malaria Ag. *P.f./P.v.* Card Test was given priority for WHO prequalification assessment.

Dossier assessment

Premier Medical Corporation Private Limited submitted a product dossier for **First Response Malaria Ag.** *P.f./P.v.* **Card Test** as per the "*Instructions for compilation of a product dossier*" (PQDx_018 v1). The information (data and documentation) submitted in the product dossier was reviewed by WHO staff and external technical experts (assessors) appointed by WHO. The manufacturer's responses to the nonconformities found during dossier screening and assessment findings were accepted on 7 September 2018.

Based on the product dossier screening and assessment findings, the product dossier for **First Response Malaria Ag.** *P.f./P.v.* **Card Test** meets WHO prequalification requirements.

Manufacturing site inspection

An onsite inspection of Premier Medical Corporation Private Limited., at A1-302 and 3704-05, GIDC, Sarigam INA, 396155 Gujarat, India, was conducted from the 22nd to 24th September 2024. At the time of considering the product application for Prequalification, the Manufacturer of the product had a well-established quality management system and manufacturing practices in place that would support the manufacture of a product of consistent quality. Routine inspections of the Manufacturing site will be conducted with copies of the WHO Public Inspection Report (WHOPIR) published on the WHO Prequalification web page as per Resolution WHA57.14 of the World Health Assembly. Note that a WHOPIR reflects the information on the most current assessment performed at a manufacturing site for in vitro diagnostic products and summarizes the assessment findings.

https://extranet.who.int/pqweb/vitro-diagnostics/who-public-inspection-reports

All published WHOPIRs are with the agreement of the manufacturer.

The onsite inspection was accepted on 10 February 2025.

Based on the site inspection and corrective action plan review, the quality management system for First Response Malaria Ag. *P.f./P.v.* Card Test meets WHO prequalification requirements.

Product performance evaluation

The eighth round of WHO product testing of RDTs for malaria antigen detection was completed in 2018. The product was evaluated against a *Plasmodium falciparum* cultured line panel, *P. falciparum* wild type parasite panel, *P. vivax* wild type parasite panel and a *Plasmodium spp.* negative panel. Thermal stability was assessed after 2 months of storage at elevated temperature and humidity, and a descriptive ease of use assessment was recorded.

Based on the demonstrated P. falciparum panel detection score (94.0% at 200 parasites/ μ l), P. vivax panel detection score (100% at 200 parasites/ μ l), false-positive rates (1.0% for clean negatives, 0.8% for P. falciparum at 200 parasites/ μ l, 0.7% for P. vivax at 200 parasites/ μ l, 0.5% for P. falciparum at 2000 to 5000 parasites/ μ l, 0.0% for P. vivax at 2000 to 5000 parasites/ μ l) and invalid rate (0.1%), First Response® Malaria Ag. P.f./P.v. Card Test meets the current laboratory evaluation requirements for prequalification.

Summary	Panel detection score (%)		False	positive	rate (%)	Invalid	
performance characteristics	200 parasites/μl		200 parasites/μl		Clean negatives	rate (%)	
	P <i>f</i>	Pv	P <i>f</i>	Pv			
First Response Malaria Ag. <i>P.f./P.v.</i> Card Test	94.0	100	0.8	0.7	1.0	0.1	

Labelling

- 1. Labels
- 2. Instructions for use

1. Labels

Packaging artwork

Alcohol swab label

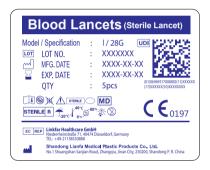






Sterile Twist lancet label

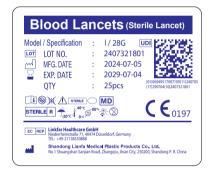
Size:50*40mm



Size:50*40mm



Size:50*42mm



Size:50*42mm





Part No:(S)PI19-CAR-008 Rev. AA 2025-01

Product Name: F.R. Malaria Ag. P.f./P.v. Card Test

Pack Size: 05 Tests / bulk

Dimension:115 (L) X 80 (W) X 40 (H) MM & GSM: 450

Language: English





Part No. : (S)PI19-CAR-005 Rev.: AC, 2025-01 Product Name : FR Malaria Ag. *P.f /P.v.* Card Test

Pack Size: 10 Tests / bulk

Dimension:115 (L) X 80 (W) X 55(H) MM & GSM: 450

Language: English





Product Name: FR Malaria Ag. P.f/P.v. Card Test

Pack Size: 25 Tests / bulk





Product Name: FR Malaria Ag. P.f/P.v. Card Test

Pack Size: 30 Tests / bulk



Card Test

(Whole Blood)

Instructions for use

Rev.: AB, 2024-10



Tel.: +91 260 2780112/113, www.premiermedcorp.com



One Step Malaria Ag. *P.f./P.v.*



Product Name: FR Malaria Ag. P.f /P.v. Card Test

Pack Size: 10 Single Tests

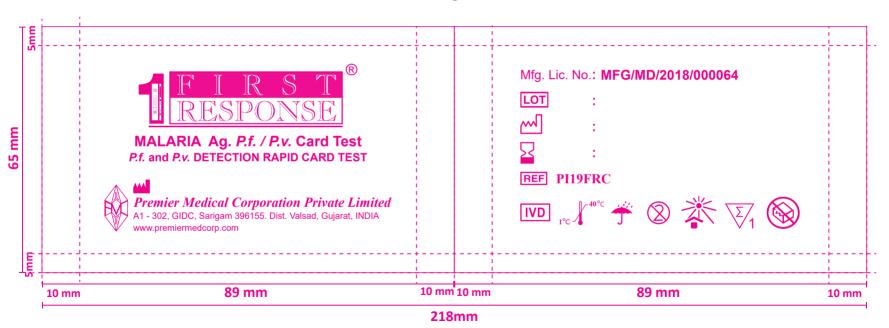




Product Name: FR Malaria Ag. *P.f /P.v.* Card Test Pack Size: 25 Single Tests



Aluminum Pouch F.R Malaria Ag. P.f / P.v Card Test- PI19FRC



Assay buffer label- First Response Malaria Ag. P.f/P.v Card Test 2.5 ml



Assay buffer label- First Response Malaria Ag. P.f/P.v Card Test 3.0 ml



2. Instructions for use¹

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 $^{^{1}}$ English version of the IFU was the one that was assessed by WHO. It is the responsibility of the manufacturer to ensure correct translation into other languages.

Instructions for Use: (Bulk Pack Size)

Specimen collection and storage

[Collection by venipuncture]

- 1) Collect the whole blood into the collection tube (containing EDTA/sodium citrate/heparin) by venipuncture.
- 2) If specimens are not immediately tested (within 1 hour) should be stored at 2-8 °C maximum upto 72 hours (3 days). Using the specimen more than three days can cause non-specific reaction

Capillary blood specimen collection:



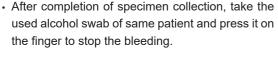
- Wear gloves, massaging the fingertip gently. It will help to obtain a round drop of blood.
- Wipe the complete finger tip with the alcohol swab. Wait until the finger tip is dried completely.

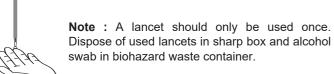


 Detach the protective cap of the lancet and pierce the end of finger tip with the sterile lancet provided.



 Gently squeeze the area until you get enough blood specimen.





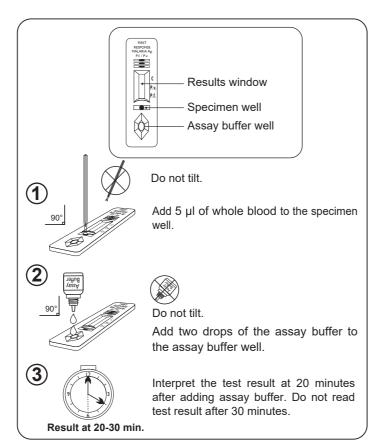
- Do not share used lancets with another person. To prevent possible infection, a used lancet should not be touched by another person.
- Do not use expired lancet. The use of an expired lancet may cause any infection at the punctured skin due to cease to exist its sterility.
- Use new lancet and choose a different puncture site, if repeat the finger prick.
- · Do not share used alcohol swab.

Specimen storage

1) Whole blood specimen may be used for testing immediately (within 1 hour) or may be stored at 2-8°C for maximum up to 72 hours (3 days). Do not use blood specimen stored for more than 3 days, it can cause non-specific reaction.

Test Procedure

- 1) Bring the First Response[®] Malaria Ag. *P.f. / P.v.* Card Test kit components to room temperature (15 40°C) prior to 15 minutes of testing.
- 2) Remove the test device and the specimen transfer device from the kit and place it on a flat, dry surface and label the test device with specimen identification number/name.
- 3) Slowly add 5 µl of whole blood to the specimen well using the specimen transfer device. Dispose the used specimen transfer device as biohazard waste immediately after use.
- 4) Add two drops of the assay buffer to the assay buffer well.
- 5) Observe for development of colored bands in the Results Window.
- Interpret test results at 20 minutes. (After recording the results, dispose of test device as a biohazardous waste).
- 7) Do not interpret after 30 minutes.



Caution

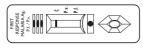
- Hold specimen transfer device and assay buffer bottle vertically, else it can lead to inaccurate results.
- Exactly 2 drops of assay buffer should be added. Adding more
 or less than 2 drops may cause over flooding or reverse
 migration phenomenon, which may lead inaccurate results of the
 test.
- Results can be interpreted any time from 20 to 30 minutes. Do not read test result after 30 minutes. Reading beyond 30 minutes may give inaccurate results. After recording the results, dispose of test device as a biohazard waste.

Internal Quality Control

The visualization of the control line in First Response® Malaria Ag. *P.f. / P.v.* Card Test indicates that active ingredient of the strips are functional and the migration is successful. The control line in First Response® Malaria Ag. *P.f. / P.v.* Card Test is not meant for specimen addition monitoring.

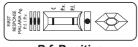
How to Interpret test results

Negative Results



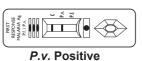
If only one color line appears, at control line 'C' as in the figure, the specimen is negative.

Positive Results



If two color lines appears, one at control line 'C' and other at test line *P.f.* as in the figure, the specimen is reactive for antigens to *P.f.*Interpret faint line as reactive line

P.f. Positive

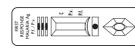


If two color lines appears, one at control line 'C' and other at test line *P.v.* as in the figure, the specimen is reactive for antigens to *P.v.*Interpret faint line as reactive line.

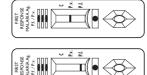


If all three color lines appears, one at control line 'C' and other two at test lines *P.f.* and *P.v.* as in the figure, the specimen is reactive for antigens to *P.f.* and *P.v.* Interpret faint line as reactive line.

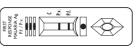
Invalid Results



No presence of control line 'C' in the result window (irrespective of presence of test lines) indicates an invalid result.



The directions may not have been followed correctly or the test may have deteriorated.



The Invalid test results should be retested with new test device.

Performance Characteristics

First Response® Malaria Ag. *P.f. / P.v.* Card Test were tested using an in-house panel of Positive and Negative clinical specimens characterized by malaria microscopy as the reference method. First Response® Malaria Ag. *P.f. / P.v.* Card Test showed 100% sensitivity and 100% specificity. First Response® Malaria Ag. *P.f. / P.v.* Card Test showed 100% agreement with the reference method.

Specimen details	Reference (Micro	e Method scopy)	First Response® Malaria Ag. P.f. / P.v. Card Test			
	Positive	Negative	Positive	Negative	Total	
P. falciparum Positive Whole blood specimen	231	0	231	0	231	
P. vivax Positive Whole blood specimen	243	0	243	0	243	
Malaria Negative Whole blood specimen	0	1287	0	1287	1287	
Total	474	1287	474	1287	1761	

Reference	Specimen details		First Response® Malaria Ag. P.f. / P.v. Card Te				
Method	Орсон	ion dotallo			Total	95% Confidence	
	Clinical Status	Parameter	Positive	Negative	Result	Interval	
Microscopy	P. falciparum Positive	Sensitivity	231	00	231	(97.96%-100%)	
Micro	P. Vivax Positive	Sensitivity	243	00	243	(98.06%-100%)	
	Malaria Negative	Specificity	00	1287	1287	(99.63%-100%)	

Worldwide Performance Panel

The analytical sensitivity of the First Response® Malaria Ag. *P.f. / P.v.* Card Test was carried out by testing WHO worldwide performance panel. Total 10 specimens were tested in-house. The First Response® Malaria Ag. *P.f. / P.v.* Card Test showed 100% Sensitivity.

Analytical Sensitivity : In- House Evaluation						
Total	Specimens	First Response® Malaria Ag. P.f./P.v. Card Test				
Total Specimens		Positive	Negative			
05	200 p/µl	05	00			
05 2000 p/µl		05	00			

Cross Reactivity Study

First Response® Malaria Ag. *P.f. / P.v.* Card Test was tested with specimen reactive for other diseases/conditions (mentioned in following table), which may interfere with performance of the test. The First Response® Malaria Ag. *P.f. / P.v.* Card Test tested with mentioned specimen for cross reactivity study as well as same diseased/condition specimens# were also used for spiking of malaria positive specimens# to determine effect on sensitivity of the test. None of the specimens interfere with the test results of First Response® Malaria Ag. *P.f. / P.v.* Card Test, and showed no cross reactivity with 100% sensitivity.

Specimens tested	P. falciparum positive specimens	P.vivax positive specimens	Malaria negative specimens	Specimens tested	P. falciparum positive specimens	P.vivax positive specimens	Malaria negative specimens
Syphilis Positive#	06	06	10	HSV 1/2 Positive#	05	05	05
HIV Positive#	03	03	05	HTLV- I Ab Positive#	07	07	07
Dengue NS1 Positive#	05	05	05	HTLV- II Ab Positive#	09	09	09
Multipara (Pregnant Woman)	08	04	72	HSV - I IgG Positive#	08	08	08
CMV Positive#	03	03	03	Rubella IgG & IgM Positive#	15	15	15
ANA Positive#	04	04	04	HBsAg Positive#	03	03	05
HAV Positive#	04	04	04	Chikungunya Positive#	05	05	05
EBV Positive#	02	02	02	Anti-malarial drug medication	02	02	Not tested
HCV Positive#	03	03	05	Anti-TB drug medication#	03	03	03
Yellow fever virus# post immunization	03	03	04	Infuenza A and B#	03	03	06
Measles [#]	03	03	04	Visceral leishmaniasis*	03	03	05
Leptospirosis#	03	03	05	Bilirubin [#]	03	03	05
Cholesterol # Triglycerides	03	03	05	Chagas	03	03	05
Influenza vaccine# recipient	03	03	05	Sickle cell	03	03	05
Leishmaniasis#	03	03	05	Acute hepatitis A infection#	03	03	05
Schistosomiasis#	03	03	05	Toxoplasmosis#	03	03	05
D. C C. I			1				

Potential interference substances

The interfering substances that may affect performance of the First Response® Malaria Ag. *P.f. / P.v.* Card Test are mentioned in following table. The First Response®Malaria Ag. *P.f. / P.v.* Card Test showed no reactivity with any of mentioned specimens# and showed 100% specificity. The same specimens# were spiked in malaria positive specimens respectively and tested.First Response® Malaria Ag. *P.f. / P.v.* Card Tests showed 100% sensitivity with spiked specimens.

Specimens tested	P. falciparum positive specimens	P.vivax positive specimens	Malaria negative specimens	Specimens tested	P. falciparum positive specimens	P.vivax positive specimens	Malaria negative specimens
Lipemic specimen#	05	05	05	Low Hematocrit specimens	Not tested	Not tested	05
Icteric specimens#	05	05	05	Whole blood specimen in ACD anticoagulant	03	03	182
Hemolytic specimens	01	01	05	RF Ab Positive#	04	04	09
High Hematocrit specimens	Not tested	Not tested	05	dsDNA Antibody Positive	01	01	01
Recipient of multiple blood transfusion	03	03	05				

Precision

- a) Within run, precision was determined by using 225 replicates of 9 different specimens containing different malaria parasitic count. Within run, precision was observed 100%.
- b) Between run, precision was determined by using the 9 different specimens containing different malaria parasitic count in 5 different replicates with 3 different lots of test devices X 5 different days X 3 different sites tested.

Between run, precision was observed 100%.

External Evaluation Report

Place of Evaluation Year Sensitivity			Specificity				
Flace of Evaluation	i cai	Р	P.f. P.v.		P.f.	P.v.	
		200 p/µl	2000 p/µl	200 p/µl 2000 p/µl			F.V.
WHO Evaluation Round 8	2018	94%	100%	100% 100%		99.0%	
Ministry of Health & Children Care Zimbabwe	2017	100%		100%		100%	100%
National Public Health & Reference Lab, Ghana	2016	100%		100%		100%	100%

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②

Potential interference drug substances

The details of interference drug molecules are mentioned in following table. Each interfering drug molecule substances were spiked at final concentration of 250µg/ml in malaria positive specimen as well as negative specimens, respectively. No false positive or false negative results were observed with any of drug molecules, when tested with First Response® Malaria Ag. P.f./P.v. Card Test

Diclofenac	Acetaminophen	Aspirin		
Folic acid	Pyrazinamide	Ampicillin Sodium salt		
Abacavir	Cholecalciferol	Nevirapine		
Magnesium sulphate	Ritonavir	Ibuprofen		
Daruvir	Rifampicin	Ascorbic Acid		
Naproxen IP	Metformin	Hydrochlorothiazide		
Pantoprazole	Isoniazid	Ferrous Ascorbate		
Ergocalciferol	Iron Chloride	Penicillin G Benzathine		
Cyclobenzaprine Hydrochloride				

Limitation

- 1) The test procedure, precautions and interpretation of results for this test must be followed while testing.
- 2) The following anticoagulants have been validated for use with this test: heparin, EDTA & citrate.
- 3) Interfering specimens like hemolytic (hemoglobin containing) samples, icteric (bilirubin containing) samples and lipemic samples do not affect the test results.
- Do not mix reagent from different lots.
- 5) Interpret faint line as a reactive line. Repeat the test in case of very faint test line or if have any doubt for test line.
- 6) Although the test is very accurate in detecting HRP2 and pLDH a low incidence of false results can occur. As with all diagnostic tests, a definitive clinical diagnosis should not be based on the results of a single test, but should only be made by the physician after all clinical and laboratory findings have been evaluated.
- 7) False negative results may arise due to very low parasite density (for instance < 100 p/µl), very high parasite density (prozone/hook effect), mutations in the HRP2 gene with deletion of HRP2 antigen, damage by heat, freezing or humidity, application of insufficient volume of blood on the device and use of wrong buffer.
- 8) False positive results can occur due to various conditions such as rheumatoid factors, antinuclear antibodies, chronic viral infection (hepatitis B or C), parasitic infection (schistosomiasis and trypanosomiasis) and use of wrong buffer.

- 1) Clinical and Laboratory Standards Institute. Procedures and devices for the collection of diagnostic capillary blood specimens; approved standard, fifth edition. CLSI H04-A6, Vol. 28, No. 25, 2008.
- 2) http://vassarstats.net/clin1.html#def , Richard Lowry.
- 3) Clinical and Laboratory Standards Institute. Procedures for the collection of diagnostic blood specimens by venipuncture: approved standard, sixth edition. CLSI H03-A6, Vol. 27, No. 26,
- 4) World Health Organization: Laboratory biosafety manual, third edition. Geneva: WHO; 2004.
- http://www.who.int/csr/resources/publications/biosafety/Biosafety7.pdf
- 5) WHO (2024). World malaria report, World Health Organization, Geneva, Switzerland. https://www.who.int/teams/global-malaria-programme/reports/world-malaria-report-2024

- Malaria rapid diagnostic test performance: results of WHO product testing of malaria RDTs: round 8 (2016-2018).
- Gillet P, Scheirlinck A, Stokx J, De Weggeleire A, Chauque H, Canhanga O. Tadeu B. Mosse C. Tiago A. Mabunda S. Bruggeman C, Bottieau E, Jacobs J: Prozone in malaria rapid diagnostics tests: how many cases are missed? Malar J 2011, 10:166. http://www.malariajournal.com/content/10/1/166
- 8) Gillet P, Mori M, Van Den Ende J, Jacobs J: Buffer substitution in malaria rapid diagnostic tests causes false positive results. Malar J 2010, 9:215 http://www.malariajournal.com/content/9/1/215
- 9) Maltha J, Gillet P, Cnops L, Van Den Ende J, Van Esbroeck M, Jacobs J: Malaria rapid diagnostic tests: Plasmodium falciparum infections with high parasite densities may generate false positive Plasmodium vivax pLDH lines. Malar J 2010, 9:198. http://www.malariajournal.com/content/9/1/198
- 10) Gamboa D, Ho M, Bendezu J, Torres K, Chiodini P, Barnwell J, Incardona S, Perkins M, Bell D, McCarthy J, Cheng Q: A large proportion of P. falciparum isolates in the Amazon region of Peru lack pfhrp2 and pfhrp3: implications for malaria rapid diagnostic tests. PLoS One 2010, 5:e8091. http://www.plosone.org/article/i fo%3Adoi%2F10.1371%2Fjournal.pone.0008091

SYMBOL L	EGENDS		
Symbol	Explanation of symbol	Symbol	Explanation of symbol
Ţi	Consult instructions for use	₹ E	Contains sufficient for < n > tests
NON	Non Sterile	REF	Product Code
IVD	In vitro diagnostic medical device	LOT	Lot Number
1°C - 40°C	Store at 1-40 °C	***	Manufacturer
\triangle	Caution	_w	Date of manufacture (YYYY-MM)
*	Keep dry		Expiration Date (YYYY-MM)
2	Do not reuse		Do not use if package is damaged
茶	Keep away from sunlight		

Product Disclaimer and Warnings

Every warnings and precautions should be taken in to consideration before using the test. Failure to consider "Precautions, Warnings and Limitations" may not ensure the diagnostic ability and accuracy of this product. The test result may accordingly be affected by environmental factors and / or user error outside of the control of the Manufacturer and Distributor.

A definitive clinical diagnosis should not be based on the results of a single test, but it should be made by physician after all clinical and laboratory findings have been evaluated.

"In no event shall our company or its distributor be liable for any direct, indirect, punitive, unforeseen, incidental, special consequential damages, to property or life, whatsoever arising out of or connected with an incorrect diagnosis, whether positive or negative, in the use or misuse of this product".

Manufactured by



Premier Medical Corporation Private Limited A1-302, GIDC, Sarigam 396155. Dist. Valsad, Gujarat, INDIA. Customer support E-mail: info@premiermedcorp.com Tel.: +91 2602780112/113 •Website: www.premiermedcorp.com

• ISO 13485 & EN ISO 13485 Certified Company

Part No.(S)PI19-INS-001, Rev.: AD, Date: 2025-09-21 ENGLISH Note: Instructions for use will be printed in local language of the country using the test, if required



FIRST RESPONSE® MALARIA Ag. P.f. / P.v. CARD TEST

A rapid test for the detection of Malarial species P. falciparum and P. vivax in human whole blood.

REF PI19FRC05, PI19FRC10, PI19FRC25, PI19FRC30



First Response® Malaria Ag. P.f./P.v. Card Test is intended to be performed by trained users (In either laboratory or point of care settings) as qualitative screening in vitro diagnostic test for detection of P. falciparum and P. vivax. The test is intended to be used with human whole blood specimens (capillary or venous blood). Venous blood with the anti-coagulants heparin, EDTA or sodium citrate do not affect the test results. This kit is not intended to be used for screening of blood donors. The test is not automated and does not require any additional instrument.

Introduction

Malaria is a serious, sometimes fatal, parasitic disease characterized by fever, chills, and anemia and is caused by four species of plasmodium parasites that is transmitted from one human to another by the bite of infected Anopheles mosquitoes. There are four Plasmodium species that can infect humans: Plasmodium falciparum, P. vivax, P. ovale and P. malariae. In humans, the parasites (called sporozoites) migrate to the liver where they mature and release another form, the merozoites into the blood which infect red blood cells. According to the latest estimates, 263 million cases of malaria occurred globally, and the diseases led to 0.6 million deaths (WHO 2024). At present, malaria is diagnosed by looking for parasites in a drop of blood.

Assay Principle

First Response® Malaria Ag. P.f. / P.v. Card Test is based on principle of immunochromatography in which nitrocellulose membrane is pre-coated with two monoclonal antibodies as two separate lines. One monoclonal antibody (test line P.v.) is P. vivax specific to lactate dehydrogenase (pLDH) and the other line (test line *P.f.*) consists of a monoclonal antibody specific to Histidine-Rich Protein 2 (HRP2) of the Plasmodium falciparum. When the test sample along with assay buffer flows through the nitrocellulose membrane, monoclonal antibodies conjugated with colloidal gold, which are P.vivax specific to pLDH and P. falciparum specific to HRP2 binds to Plasmodium antigens released from the lysed blood sample. These antigen-conjugated antibody complex moves through the nitrocellulose membrane and binds to corresponding immobilised antibody at test lines, which leads to the formation of colour line / lines indicating reactive results. The control line will appear irrespective of reactive or non reactive sample.

So, the First Response[®] Malaria Ag. P.f. / P.v. Card Test is "of additional value" in the differential diagnosis of Plasmodium falciparum and P. vivax.

Materials Provided



test device Assay buffer



Specimen transfer device

Alcohol Swab

Sterile Lancet

PI19FRC30 **Materials Provided** PI19FRC05 PI19FRC10 PI19FRC25 Test Device Pouch Containing: 1 test Device, 1 desiccant 05 Nos 10 Nos 25 Nos 30 Nos Specimen transfer device 05 Nos. 10 Nos. 25 Nos. 30 Nos. Assay buffer bottle 1 No. 1 No. 1 No. 1 No. Sterile lancet 05 Nos. 10 Nos 25 Nos. 30 Nos. Alcohol swab 05 Nos 10 Nos. 25 Nos 30 Nos Instructions for use 1 No. 1 No. 1 No. 1 No.

bottle

Materials Required but Not Provided

- New pair of disposable gloves.
- Disposable face mask, permanent marker pen and timer.
- · Extra lancets and alcohol swabs, if needed.
- Sharp disposable box and biohazardous waste container.
- Venipuncture blood collection kit (if whole blood is collected by venipuncture).

Storage and Stability

- 1) First Response® Malaria Ag. P.f. / P.v. Card Test should be stored at 1°C-40°C.
- 2) Do not freeze the kit or components.
- 3) Assay buffer (opened and unopened) and the unopened test device are stable until the expiry date printed on the label, when stored at 1°C - 40°C.
- 4) Test device is sensitive to humidity and heat if remained opened for longer period hence perform the test immediately after removing the test device from the foil pouch.
- 5) The shelf life of the kit is as indicated on the outer package.

Precautions

- 1) Wear protective gloves and face mask while handling specimens.
- 2) Dispose of used gloves as biohazard waste. Wash hands thoroughly afterwards.
- 3) Avoid splashing or aerosol formation.
- 4) Clean up spills thoroughly using an appropriate disinfectant.
- 5) Decontaminate and dispose of all used specimens, test devices, alcohol swabs and specimen transfer device as an infectious waste, in a biohazardous waste container. Dispose of used lancets in a sharp box and face mask in waste container.

Warnings

- 1) For in vitro diagnostic use only.
- 2) Read the instructions carefully before performing the test, deviation will invalidate the test results.
- 3) Apply standard biosafety precautions for handling and disposal of potentially infective material.
- 4) Assay buffer contains sodium azide as preservative which may be toxic if ingested. When disposed of through sink, flush with large quantity of water.
- 5) Devices and assay buffer of different lot must not be used.
- 6) Do not use the test device if the pouch is not intact.
- 7) Do not use the lancet if the seal is broken.
- 8) Do not use the test device if the dessicant is missing or if found saturated (orange colour has turned green).
- 9) Do not smoke, eat or drink while handling specimens and performing a test.
- 10) Do not re-use the test device, alcohol swab, lancet and specimen transfer device as are intended for single use only. 11) Perform the test by using kit assay buffer, any other buffer or
- fluid will invalidate the test results. 12) Do not allow the tip of assay buffer bottle to touch specimen
- well, it contaminate assay buffer. 13) Do not use test device and assay buffer beyond the date of
- 14) Do not eat the dessicant.

expiry.

15) Do not use any other specimen other than human whole blood & do not mix and interchange different specimens.





Instructions for Use Single test Pack Size: (Master)

Specimen collection and storage

[Collection by venipuncture]

- 1) Collect the whole blood into the collection tube (containing EDTA/sodium citrate/heparin) by venipuncture.
- 2) If specimens are not immediately tested (within 1 hour) should be stored at 2-8 °C maximum upto 72 hours (3 days). Using the specimen more than three days can cause non-specific reaction.

Capillary blood specimen collection:



- Wear gloves, massaging the fingertip gently. It will help to obtain a round drop of blood.
- Wipe the complete finger tip with the alcohol swab. Wait untill the finger tip dried completely.



 Detach the protective cap of the lancet and pierce the end of finger tip with the sterile lancet provided.

Gently squeeze the area until you get enough blood specimen.



 Take the specimen transfer device provided. Hold it vertically and gently touch the open concave end into the blood drop.



 After completion of specimen collection, take the used alcohol swab of same patient and press it on the finger to stop the bleeding.

Note: A lancet should only be used once. Dispose of used lancets in sharp box and alcohol swab in biohazard waste container.

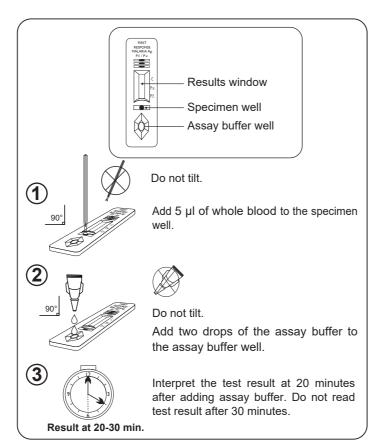
- Do not share used lancets with another person. To prevent possible infection, a used lancet should not be touched by another person.
- Do not use expired lancet. The use of an expired lancet may cause any infection at the punctured skin due to cease to exist its sterility.
- Use new lancet and choose a different puncture site, if repeat the finger prick.
- · Do not share used alcohol swab.

Specimen storage

1) Whole blood specimen may be used for testing immediately (within 1 hour) or may be stored at 2-8°C for maximum up to 72 hours (3 days). Do not use blood specimen stored for more than 3 days, It can cause non-sepcific reaction.

Test Procedure

- 1) Bring the First Response[®] Malaria Ag. *P.f. / P.v.* Card Test kit components to room temperature (15 40°C) prior to 15 minutes of testing.
- 2) Remove the test device and the specimen transfer device from the kit and place it on a flat, dry surface and label the test device with specimen identification number/name.
- 3) Slowly add 5 µl of whole blood to the specimen well using the specimen transfer device. Dispose the used specimen transfer device as biohazard waste immediately after use.
- 4) Add two drops of the assay buffer to the assay buffer well.
- 5) Observe for development of colored bands in the Results Window.
- Interpret test results at 20 minutes. (After recording the results, dispose of test device as a biohazardous waste).
- 7) Do not interpret after 30 minutes.



Caution

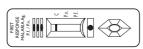
- Hold specimen transfer device and assay buffer vial vertically, else it can lead to inaccurate results.
- Exactly 2 drops of assay buffer should be added. Adding more
 or less than 2 drops may cause over flooding or reverse
 migration phenomenon, which may lead inaccurate results of
 the test.
- Results can be interpreted any time from 20 to 30 minutes.Do not read test result after 30 minutes. Reading beyond 30 minutes may give inaccurate results. After recording the results, dispose of test device as a biohazard waste.

Internal Quality Control

The visualization of the control line in First Response® Malaria Ag. *P.f. / P.v.* Card Test indicates that active ingredient of the strips are functional and the migration is successful. The control line in First Response® Malaria Ag. *P.f. / P.v.* Card Test is not meant for specimen addition monitoring.

How to Interpret test results

Negative Results



If only one color line appear, at control line 'C' as in the figure, the specimen is negative.

Positive Results



If two color lines appears, one at control line 'C' and other at test line *P.f.* as in the figure, the specimen is reactive for antigens to *P.f.*

Interprete faint line as reactive line



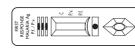
If two color lines appears, one at control line 'C' and other at test line *P.v.* as in the figure, the specimen is reactive for antigens to *P.v.*Interprete faint line as reactive line.

If a

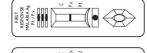
P.f. & P.v. Positive

If all three color lines appears, one at control line 'C' and other two at test lines *P.f.* and *P.v.* as in the figure, the specimen is reactive for antigens to *P.f.* and *P.v.* Interprete faint line as reactive line.

Invalid Results

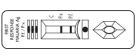


No presence of control line 'C' in the result window (irrespective of presense of test lines) indicates an invalid result.



Special Specia

The directions may not have been followed correctly or the test may have detoriorated.



The Invalid test results should be retested with new test device.

Performance Characteristics

First Response®Malaria Ag. *P.f. / P.v.* Card Test were tested using an in-house panel of Positive and Negative clinical specimens characterized by malaria microscopy as the reference method. First Response® Malaria Ag. *P.f. / P.v.* Card Test showed 100% sensitivity and 100% specificity. First Response® Malaria Ag. *P.f. / P.v.* Card Test showed 100% agreement with the reference method.

Specimen details	Reference Method (Microscopy)		First Response® Malaria Ag. P.f. / P.v. Card Test			
	Positive	Negative	Positive	Negative	Total	
P. falciparum Positive Whole blood specimen	231	0	231	0	231	
P. vivax Positive Whole blood specimen	243	0	243	0	243	
Malaria Negative Whole blood specimen	0	1287	0	1287	1287	
Total	474	1287	474	1287	1761	

Reference	Specimen details		Specimen details First Response® Malaria Ag. P.f. / P.v. Card			
Method	Clinical Status	Parameter	Positive	Negative	Total Result	95% Confidence Interval
Microscopy	P. falciparum Positive	Sensitivity	231	00	231	(97.96%-100%)
Micro	P. Vivax Positive	Sensitivity	243	00	243	(98.06%-100%)
	Malaria Negative	Specificity	00	1287	1287	(99.63%-100%)

Worldwide Performance Panel

The analytical sensitivity of the First Response® Malaria Ag. *P.f. / P.v.* Card Test was carried out by testing WHO worldwide performance panel. Total 10 specimens were tested in-house. The First Response® Malaria Ag. *P.f. / P.v.* Card Test showed 100% Sensitivity.

Analytical Sensitivity : In- House Evaluation					
Total	Specimens	First Response® Malari	a Ag. P.f./P.v. Card Test		
Iotal	Specimens	Positive	Negative		
05	200 p/µl	05	00		
05	2000 p/µl	05	00		

Cross Reactivity Study

First Response® Malaria Ag. *P.f. / P.v.* Card Test was tested with specimen reactive for other diseases/conditions (mentioned in following table), which may interfere with performance of the test. The First Response® Malaria Ag. *P.f. / P.v.* Card Test tested with mentioned specimen for cross reactivity study as well as same diseased/condition specimens# were also used for spiking of malaria positive specimens# to determine effect on sensitivity of the test. None of the specimens interfere with the test results of First Response® Malaria Ag. *P.f. / P.v.* Card Test, and showed no cross reactivity with 100% sensitivity.

ecimens tested	P. falciparum positive specimens	P.vivax positive specimens	Malaria negative specimens	Specimens tested	P. falciparum positive specimens	P.vivax positive specimens	Malaria negative specimens
yphilis Positive#	06	06	10	HSV 1/2 Positive#	05	05	05
HIV Positive#	03	03	05	HTLV- I Ab Positive#	07	07	07
Dengue NS1 Positive#	05	05	05	HTLV- II Ab Positive#	09	09	09
Multipara regnant Woman)	08	04	72	HSV - I IgG Positive#	08	08	08
CMV Positive#	03	03	03	Rubella IgG & IgM Positive#	15	15	15
ANA Positive#	04	04	04	HBsAg Positive#	03	03	05
HAV Positive#	04	04	04	Chikungunya Positive#	05	05	05
EBV Positive#	02	02	02	Anti-malarial drug medication	02	02	Not tested
HCV Positive#	03	03	05	Anti-TB drug medication#	03	03	03
ellow fever virus# st immunization	03	03	04	Infuenza A and B#	03	03	06
Measles [#]	03	03	04	Visceral leishmaniasis*	03	03	05
Leptospirosis [#]	03	03	05	Bilirubin [#]	03	03	05
Cholesterol # Triglycerides	03	03	05	Chagas	03	03	05
fluenza vaccine# recipient	03	03	05	Sickle cell	03	03	05
Leishmaniasis#	03	03	05	Acute hepatitis A infection#	03	03	05
chistosomiasis#	03	03	05	Toxoplasmosis#	03	03	05

Potential interference substances

The interfering substances that may affect performance of the First Response® Malaria Ag. *P.f. / P.v.* Card Test are mentioned in following table. The First Response®Malaria Ag. *P.f. / P.v.* Card Test showed no reactivity with any of mentioned specimens* and showed 100% specificity. The same specimens* were spiked in malaria positive specimens respectively and tested.First Response® Malaria Ag. *P.f. / P.v.* Card Tests showed 100% sensitivity with spiked specimens.

Specimens tested	P. falciparum positive specimens	P.vivax positive specimens	Malaria negative specimens	Specimens tested	P. falciparum positive specimens	P.vivax positive specimens	Malaria negative specimens
Lipemic specimen#	05	05	05	Low Hematocrit specimens	Not tested	Not tested	05
Icteric specimens#	05	05	05	Whole blood specimen in ACD anticoagulant	03	03	182
Hemolytic specimens	01	01	05	RF Ab Positive#	04	04	09
High Hematocrit specimens	Not tested	Not tested	05	dsDNA Antibody Positive	01	01	01
Recipient of multiple blood transfusion	03	03	05				_

Precision

- a) Within run, precision was determined by using 225 replicates of 9 different specimens containing different malaria parasitic count. Within run, precision was observed 100%.
- b) Between run, precision was determined by using the 9 different specimens containing different malaria parasitic count in 5 different replicates with 3 different lots of test devices X 5 different days X 3 different sites tested.

Between run, precision was observed 100%

External Evaluation Report

Place of Evaluation	Year	Sensitivity			Specificity		
Place of Evaluation	Teal	P.f. P.v.		P.f.	P.v.		
		200 p/µl	2000 p/µl	200 p/µl	2000 p/µl	Г.І.	Γ.V.
WHO Evaluation Round 8	2018	94% 100% 100% 100%		99.0%			
Ministry of Health & Children Care Zimbabwe	2017	100%		10	0%	100%	100%
National Public Health & Reference Lab, Ghana	2016	100%		100%		100%	100%



2

Potential interference drug substances

The details of interference drug molecules are mentioned in following table. Each interfering drug molecule substances were spiked at final concentration of 250µg/ml in malaria positive specimen as well as negative specimens, respectively. No false positive or false negative results were observed with any of drug molecules, when tested with First Response® Malaria Ag. P.f./P.v. Card Test

Diclofenac	Acetaminophen	Aspirin		
Folic acid	Pyrazinamide	Ampicillin Sodium salt		
Abacavir	Cholecalciferol Nevirapine			
Magnesium sulphate	Ritonavir	Ibuprofen		
Daruvir	Rifampicin	Ascorbic Acid		
Naproxen IP	Metformin	Hydrochlorothiazide		
Pantoprazole	Isoniazid	Ferrous Ascorbate		
Ergocalciferol	Iron Chloride	Penicillin G Benzathine		
Cyclobenzaprine Hydrochloride				

Limitation

- 1) The test procedure, precautions and interpretation of results for this test must be followed while testing.
- 2) The following anticoagulants have been validated for use with this test: heparin, EDTA & citrate.
- 3) Interfering specimens like hemolytic (hemoglobin containing) samples, icteric (bilirubin containing) samples and lipemic samples do not affect the test results.
- 4) Do not mix reagent from different lots.
- 5) Interpret faint line as a reactive line. Repeat the test in case of very faint test line or if have any doubt for test line.
- 6) Although the test is very accurate in detecting HRP2 and pLDH a low incidence of false results can occur. As with all diagnostic tests, a definitive clinical diagnosis should not be based on the results of a single test, but should only be made by the physician after all clinical and laboratory findings have been evaluated.
- 7) False negative results may arise due to very low parasite density (for instance < 100 p/µl), very high parasite density (prozone/hook effect), mutations in the HRP2 gene with deletion of HRP2 antigen, damage by heat, freezing or humidity, application of insufficient volume of blood on the device and use of wrong buffer.
- 8) False positive results can occur due to various conditions such as rheumatoid factors, antinuclear antibodies, chronic viral infection (hepatitis B or C), parasitic infection (schistosomiasis and trypanosomiasis) and use of wrong buffer.

References

- 1) Clinical and Laboratory Standards Institute. Procedures and devices for the collection of diagnostic capillary blood specimens; approved standard, fifth edition. CLSI H04-A6, Vol. 28, No. 25, 2008
- 2) http://vassarstats.net/clin1.html#def, Richard Lowry.
- 3) Clinical and Laboratory Standards Institute. Procedures for the collection of diagnostic blood specimens by venipuncture; approved standard, sixth edition. CLSI H03-A6, Vol. 27, No. 26,
- 4) World Health Organization: Laboratory biosafety manual, third edition. Geneva: WHO; 2004. http://www.who.int/csr/resources/publications/biosafety/Biosafetv7.pdf
- 5) WHO (2024). World malaria report, World Health Organization, Geneva, Switzerland. https://www.who.int/teams/global-malaria-programme/reports/world-malaria-report-2024
- 6) Malaria rapid diagnostic test performance: results of WHO product testing of malaria RDTs: round 8 (2016-2018).

- Gillet P, Scheirlinck A, Stokx J, De Weggeleire A, Chauque H, Canhanga O, Tadeu B, Mosse C, Tiago A, Mabunda S, Bruggeman C, Bottieau E, Jacobs J: Prozone in malaria rapid diagnostics tests: how many cases are missed? Malar J 2011, 10:166. http://www.malariaiournal.com/content/10/1/166
- Gillet P. Mori M. Van Den Ende J. Jacobs J: Buffer substitution in malaria rapid diagnostic tests causes false positive results. Malar J 2010, 9:215 http://www.malariajournal.com/con-
- 9) Maltha J, Gillet P, Cnops L, Van Den Ende J, Van Esbroeck M, Jacobs J: Malaria rapid diagnostic tests: Plasmodium falciparum infections with high parasite densities may generate false positive Plasmodium vivax pLDH lines. Malar J 2010, 9:198. http://www.malariajournal.com/content/9/1/198
- 10) Gamboa D, Ho M, Bendezu J, Torres K, Chiodini P, Barnwell J, Incardona S, Perkins M, Bell D, McCarthy J, Cheng Q: A large proportion of P. falciparum isolates in the Amazon region of Peru lack pfhrp2 and pfhrp3: implications for malaria rapid diagnostic tests. PLoS One 2010, 5:e8091.

http://www.plosone.org/article/i fo%3Adoi%2F10.1371%2Fjournal.pone.0008091

Symbol	Explanation of symbol	Symbol	Explanation of symbol
[]i	Consult instructions for use	Σ	Contains sufficient for < n > tests
NON INTERRE	Non Sterile	REF	Product Code
IVD	In vitro diagnostic medical device	LOT	Lot Number
1°C 1 40°C	Store at 1-40 °C	***	Manufacturer
\triangle	Caution	~~	Date of manufacture (YYYY-MM)
*	Keep dry	₽	Expiration Date (YYYY-MM)
2	Do not reuse	®	Do not use if package is damaged
类	Keep away from sunlight		_

Product Disclaimer and Warnings

Every warnings and precaution should be taken in to consideration before using the test. Failure to consider "Precaution, Warning and Limitations" may not ensure the diagnostic ability and accuracy of this product. The test result may accordingly be affected by environmental factors and / or user error outside of the control of the Manufacturer and Distributor.

A definitive clinical diagnosis should not be based on the results of a single test, but it should be made by physician after all clinical and laboratory findings have been evaluated.

"In no event shall our company or its distributor be liable for any direct, indirect, punitive, unforeseen, incidental, special consequential damages, to property or life, whatsoever arising out of or connected with an incorrect diagnosis, whether positive or negative, in the use or misuse of this product".

Manufactured by

Premier Medical Corporation Private Limited A1-302, GIDC, Sarigam 396155. Dist. Valsad, Gujarat, INDIA. Customer support E-mail: info@premiermedcorp.com Tel.: +91 2602780112/113 •Website: www.premiermedcorp.com

• ISO 13485 & EN ISO 13485 Certified Company

Part No.(S)PI19-INS-003, Rev.: AD Date: 2025-09-21 ENGLISH Note: Instructions for use will be printed in local language of the country using the test, if required



FIRST RESPONSE® MALARIA Ag. P.f. / P.v. CARD TEST

A rapid test for the detection of Malarial species P. falciparum and P. vivax in human whole blood.

REF PI19FRC10s & PI19FRC25s



Intended Use

First Response® Malaria Ag. P.f. / P.v. Card Tests intended to be performed by trained users (In either laboratory or point of care settings) as qualitative screening in vitro diagnostic test for detection of P. falciparum and P. vivax. The test is intended to be used with human whole blood specimens (capillary or venous blood). Venous blood with the anti-coagulants heparin, EDTA or sodium citrate do not affect the test results. This kit is not intended to be used for screening of blood donors. The test is not automated and does not require any additional instrument.

Introduction

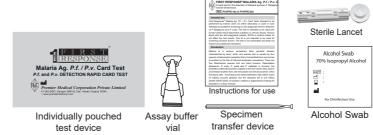
Malaria is a serious, sometimes fatal, parasitic disease characterized by fever, chills, and anemia and is caused by four species of plasmodium parasites that is transmitted from one human to another by the bite of infected Anopheles mosquitoes. There are four Plasmodium species that can infect humans: Plasmodium falciparum, P. vivax, P. ovale and P. malariae. In humans, the parasites (called sporozoites) migrate to the liver where they mature and release another form, the merozoites into the blood which infect red blood cells. According to the latest estimates, 263 million cases of malaria occured globally, and the diseases led to 0.6 million deaths (WHO 2024). At present, malaria is diagnosed by looking for parasites in a drop of blood.

Assay Principle

First Response® Malaria Ag. P.f. / P.v. Card Test is based on principle of immunochromatography in which nitrocellulose membrane is pre-coated with two monoclonal antibodies as two separate lines. One monoclonal antibody (test line P.v.) is P. vivax specific to lactate dehydrogenase (pLDH) and the other line (test line *P.f.*) consists of a monoclonal antibody specific to Histidine-Rich Protein 2 (HRP2) of the Plasmodium falciparum. When the test sample along with assay buffer flows through the nitrocellulose membrane, monoclonal antibodies conjugated with colloidal gold, which are P.vivax specific to pLDH and P. falciparum specific to HRP2 binds to Plasmodium antigens released from the lysed blood sample. These antigen-conjugated antibody complex moves through the nitrocellulose membrane and binds to corresponding immobilised antibody at test lines, which leads to the formation of colour line / lines indicating reactive results. The control line will appear irrespective of reactive or non reactive sample.

So, the First Response® Malaria Ag. P.f. / P.v. Card Test is "of additional value" in the differential diagnosis of Plasmodium falciparum and P. vivax.

Materials Provided



Materials Provided	PI19FRC10s	PI19FRC25s
Each single test pack contents: (Test device with desiccant, specimen transfer device, alcohol swab, sterile lancet, buffer vial & condensed instructions for use)	10 Nos.	25 Nos.
Master instructions for use	1 No.	1 No.

Materials Required but Not Provided

- New pair of disposable gloves.
- Disposable face mask, permanant marker pen and timer.
- · Extra lancets and alcohol swabs, if needed.
- Sharp disposable box and biohazardous waste container.
- Venipuncture blood collection kit (if whole blood is collected by venipuncture).

Storage and Stability

- 1) First Response® Malaria Ag. P.f. / P.v. Card Test should be stored at 1°C-40°C.
- Do not freeze the kit or components.
- Unopened assay buffer vial and test device are stable until the expiry date printed on the kit, when stored at 1°C - 40°C.
- Test device is sensitive to humidity and heat if remained opened for longer period hence perform the test immediately after removing the test device from the foil pouch.
- 5) The shelf life of the kit is as indicated on the outer package.

Precautions

- 1) Wear protective gloves and face mask while handling specimens.
- 2) Dispose of used gloves as biohazard waste. Wash hands thoroughly afterwards.
- 3) Avoid splashing or aerosol formation.
- 4) Clean up spills thoroughly using an appropriate disinfectant.
- 5) Decontaminate and dispose of all used specimens, test devices, alcohol swabs and specimen transfer device as an infectious waste, in a biohazardous waste container. Dispose of used lancets in a sharp box and face mask in waste container.

Warnings

- 1) For in vitro diagnostic use only.
- 2) Read the instructions carefully before performing the test, any deviation will invalidate the test results.
- 3) Apply standard biosafety precautions for handling and disposal of potentially infective material.
- 4) Assay buffer contains sodium azide as preservative which may be toxic if ingested. When disposed of through sink, flush with large quantity of water.
- 5) Devices and assay buffer of different lot must not be used.
- 6) Do not use the test device if the pouch is not intact.
- 7) Do not use the lancet if the seal is broken.
- 8) Do not use the test device if the desiccant is missing or if found saturated (orange colour has turned green).
- 9) Do not smoke, eat or drink while handling specimens and performing a test.
- 10) Do not re-use the test device, alcohol swab, assay buffer vial, lancet and specimen transfer device as are intended for single use only.
- 11) Perform the test by using kit assay buffer, any other buffer or fluid will invalidate the test results.
- 12) Do not use test device and assay buffer beyond the date of expiry.
- 13) Do not eat the desiccant.
- 14) Do not use any other specimen other than human whole blood & do not mix and interchange different specimens.



Instructions for Use Single test Pack Size: (Condensed)



First Response® Malaria Ag. P.f. / P.v. Card Test

A rapid test for the detection of Malarial species P. falciparum and P. vivax in human whole blood



Intended Use :

First Response® Malaria Aq. Pf. / P.v. Card Tests intended to be performed by trained users (In either laboratory or point of care settings) as qualitative screening in vitro diagnostic test for detection of P. falciparum and P. vivax. The test is intended to be used with human whole blood specimens (capillary or venous blood). Venous blood with the anti-coagulants heparin, EDTA or sodium citrate do not affect the test results. This kit is not intended to be used for screening of blood donors. The test is not automated and does not required any additional instrument.

Materials provided:

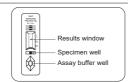
Test device with desiccant, specimen transfer device, assay buffer vial, sterile lancet, alcohol swab & instructions for use

Storage & stability:

- First Response[®] Malaria Aq. P.f. / P.v. Card Tests should be stored at 1°C - 40°C
- 2) Test device is sensitive to humidity and heat if remained opened for longer period hence perform the test immediately after removing the test device from the foil pouch.
- The shelf life of the kit is as indicated on the outer package.

Precautions & warnings:

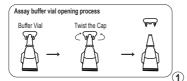
- Wear protective gloves while handling specimens.
- 2) Apply standard biosafety precautions for handling and disposal of potentially infective material
- 3) Assay buffer contains sodium azide as preservative which may be toxic if ingested. When disposed of through sink, flush with large quantity of water.
- Do not use the test device if the pouch is not intact.
- Do not use the lancet if the seal is broken.
- 6) Do not use the test device if the desiccant is missing or if found saturated (orange colour has turned green).
- 7) Do not re-use the test device, alcohol swab, lancet and specimen transfer device as are intended for single use only.
- Perform the test by using kit assay buffer, any other buffer or fluid will invalidate the test results.
- 9) Do not eat the desiccant.
- 10) Do not use any other specimen other than human whole blood & do not mix and interchange different specimens.

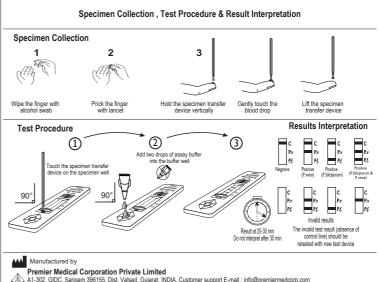


Test Procedure :

- 1) Bring the First Response® Malaria Ag. P.f. / P.v. Card Tests components to room temperature (15 - 40°C) prior to 15 minutes of
- 2) Remove the test device and the specimen transfer device from the kit and place it on a flat, dry surface and label the test device with specimen identification number/name.
- 3) Slowly add 5 µl of whole blood to the specimen well using the specimen transfer device. Dispose the used Specimen transfer device as biohazardous waste immediately after use.
- 4) Add two drops of the assay buffer to the assay buffer well.
- Observe for development of colored bands in the Results Window. 6) Interpret test results at 20 minutes. (After recording the results,
- dispose of test device as a biohazardous waste).

Do not interpret after 30 minutes.





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